REVIEW

Aspects related to oxidative stress-mediated toxicity of doxorubicin during chemotherapy treatment

Fernanda Carolina de Campos¹, Carolina Panis^{2,6}, Tatiane De Rossi³, Vanessa Jacob Victorino⁴, Alessandra Lourenço Cecchini⁵, Rubens Cecchini⁵

ABSTRACT

Objective: This study aimed to describe the main toxic effects mediated by oxidative stress associated with treatment with doxorubicin in scientific research articles available in the literature. **Material and Methods:** This study employed a descriptive review methodology applied to the literature. For the theoretical scientific background, we used the electronic PubMed search engines. **Conclusion:** The toxicity of chemotherapy treatment with doxorubicin causes damage in various organs of patients who are in uninterrupted treatment with this antineoplastic agent. Anthracycline-induced cardiotoxicity has been investigated to a great degree and is especially indicated as the principal side effect. Therefore, care needs to be given to other damage caused by this medication as important as myocardial toxicity, such as renal, pulmonary and liver toxicity, among others. There is a need for further studies to prevent or even encounter a way to control the damage caused by these toxicities in various tissues.

Keywords: hemotherapy, doxorubicin, oxidative stress, side effects, toxicity.

INTRODUCTION

Chemotherapeutic agents are important for the treatment of tumors, but since they do not discriminate healthy cells from malignant, they are toxic to all cells in division, with the accumulation of these agents occurring in healthy tissues causing serious clinical toxicity ¹. Several side effects originating from treatments with chemotherapeutic agents have been described, most associated with the interaction of the oxidative metabolism of antineoplastic agents with blood cells².

Doxorubicin (DOX) is an anthracycline drug used to treat a variety of cancers³, and it is already clear that

⁶ Stem Cell Laboratory, Brazilian National Cancer Institute (INCA), Rio de Janeiro, Brazil.

Send correspondence to:

Fernanda Carolina de Campos.

Laboratory of Pathophysiology and Free Radicals. Department of General Pathology -Biological Science Center.

Universidade Estadual de Londrina. Londrina - PR, Brasil. CEP: 86051-990. Tel: +55 (43) 3371-4521. Fax: +55 (43) 3371-4267. E-mail: fecarolina_c@hotmail.com

Submitted: 17/11/2011. Aproved: 21/09/2012. its mechanism of action consists of interference with the DNA of the cancer cell, mainly through interactions with DNA topoisomerase II^4 .

In relation to both anticarcinogenic and toxicity effects, it is widely accepted that oxidative stress and the production of free radicals are involved in DOX toxicity¹. The oxidative stress status of a cell is characterized by excessive production of reactive oxygen species (ROS) and/or a reduction in antioxidant defenses causing an imbalance in the normal metabolism of oxygen of an organism⁵. In spite of DOX being one of the most widely used anticancer agents, its use is limited due to severe damage to tissues such as heart, kidney, lung, liver and skeletal muscle¹.

Chemotherapy with DOX has, among other things, the characteristic of inducing acute vascular toxicity reducing gonadal blood volume and blood flow and femoral artery blood flow, compromising the blood vessel wall which may cause cardiovascular complications and the long-term progression of diseases such as atherosclerosis. To clarify this mechanism of vascular toxicity may be essential for the discovery of biological keys necessary to reduce the possible complications caused by treatment with DOX in survivors of cancer⁶.

Within this context, the aim of this study was to describe the principal toxic effects associated with oxidative stress during treatment with DOX in scientific research articles available in the literature.

¹ Specialist, Laboratory of Pathophysiology and Free Radicals, Universidade Estadual de Londrina, Londrina-PR, Brazil.

² Dr, Laboratory of Pathophysiology and Free Radicals, Universidade Estadual de Londrina, Londrina-PR, Brazil.

³ MSc, Laboratory of Pathophysiology and Free Radicals, Universidade Estadual de Londrina, Londrina-PR, Brazil.

⁴ Graduate Student, Laboratory of Pathophysiology and Free Radicals, Universidade Estadual de Londrina, Londrina-PR, Brazil.

⁵ PhD, Laboratory of Pathophysiology and Free Radicals, Universidade Estadual de Londrina, Londrina-PR, Brazil.

MATERIAL AND METHODS

This study employed a descriptive review methodology applied to the literature. For the theoretical scientific background, we used the electronic PubMed search engines (http://www.ncbi.nlm.nih.gov/pubmed/). Using the keywords "doxorubicin", we found 45,151 articles published during the period of 1971 until September 2011, with 3,575 review studies. Using the combination of keywords "adverse effects, doxorubicin, oxidative stress" showed 3,040 results, where the first article was published in 1972, with 948 reviews. The articles found were published in English and Portuguese, including original articles and reviews. The articles selected to compose this systematic review were in accordance with their relevance and human and experimental application, using the most recent results.

Pharmacology of doxorubicin and the generation of free radicals

Chemically, DOX is 5,12-Naphthacenedione, 10-((3-amino-2,3,6-trideoxy- α -L-ribo-hexopyranosyl)oxy)-7,8,9,10-tetrahydro-6,8,11-trihydroxy-8-(hydroxyacetyl)-1--methoxy-, hydrochloride, (8S-*cis*). DOX is a red-orange crystalline powder, soluble in water, belonging to the class of anthracyclines, a group of antineoplastic drugs discovered in the 1960s that corresponds to one of the most effective chemotherapeutic agents ever developed against cancer⁷.

The chemical structure of DOX (Figure 1) consists of a tetracyclic ring with adjacent quinone-hydroquinone groups in rings B and C, a methoxy substituent at C-4 in ring D and a short side chain at C-9 with a carbonyl at C-13. The sequence of rings A, B, C and D corresponds to the anthraquinone nucleus of anthracyclines. To date, DOX has been widely used in the treatment protocols of breast cancers, childhood solid tumors, soft tissue sarcomas and aggressive lymphomas⁸.

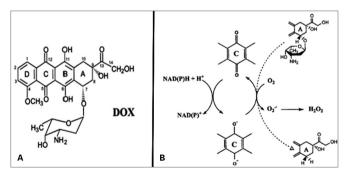


Figure 1. A: Chemical structure of doxorubicin; B: redox cycle Source: Minotti et al.⁸.

The mechanisms of action of DOX as an antineoplastic agent include intercalation into DNA leading to inhibited synthesis of macromolecules, generation of free radicals with damage to DNA and lipid peroxidation, DNA alkylation, DNA cross-linking, separation of the DNA helices, direct effects on the cell membrane altering its fluidity, topoisomerase II inhibition and induction of apoptosis depending on the concentration that chemotherapy is able to reach the site of action⁹. It has also been described as an immunomodulatory mechanism consisting of its ability to trigger the process of caspase-dependent cell death in tumor cell lines¹⁰.

The addition of one electron to the quinone moiety of ring C of DOX results in the formation of a semiquinone form that quickly regenerates the quinone form, by reducing molecular oxygen to ROS, such as hydrogen peroxide and superoxide anion in the presence of NAD(P) H-oxidoreductases (cytochrome P450, mitochondrial NADH dehydrogenase, xanthine dehydrogenase and endothelial nitric oxide synthase). The redox cycle of DOX is still accompanied by the release of iron ions from intracellular stores, resulting in the reaction of the chemotherapeutic agent with the metal ions released (in a 3:1 ratio) and interaction with hydrogen peroxide and the formation of hydroxyl radicals¹¹⁻¹³.

Such evidence has been supported by a series of studies that demonstrated the formation of free radicals during exposure to DOX in rat glioblastoma cells of, human cervical adenocarcinoma cells, mammary culture lines MCF-7 and acute lymphocytic leukemia murine cells^{9,14,15}. However, this cytotoxic mechanism has been challenged¹⁶.

Pharmacokinetic studies performed in patients with different types of tumors have shown that DOX presents a multiphase profile after intravenous injection. After intravenous administration of the usual dose of 60 to 75 mg/m², the initial half-life of distribution is approximately 5 minutes with a terminal half-life of elimination of 20 to 48 hours. It features extensive tissue binding and its main active metabolite, doxorubicinol, has plasma protein binding of approximately 70%. Elimination is predominantly via biliary excretion and oxidative metabolism¹⁷.

Due to its electron configuration, free radicals present as unstable molecules, short half-life and high reactivity with components of biological systems such as lipids, proteins and DNA¹⁸. As a consequence of the action of radicals in these structures, there are the phenomena of lipid peroxidation, protein carbonilation and DNA oxidation.

Adverse effects of doxorubicin treatment on humans

Since the 1970s, cardiotoxicity resulting from use of DOX has been described¹⁹. However, this adverse effect is only one of many presented by this anthracycline. These effects are described in the literature as pulmonary toxicity²⁰, hepatotoxic and nephrotoxic effects²¹, suggesting systemic damage not only located in the tumor.

DOX is converted into its semiquinone form within the cardiomyocytes by the P450 system and flavin-containing monooxygenase²². Experimental studies suggest that one of the mechanisms by which cardiomyocytes target the action of DOX is by reducing the levels of catalase, superoxide dismutase (SOD) and glutathione peroxidase in cardiomyocytes after chemotherapy by promoting superoxide anion- and hydrogen peroxide- induced apoptosis²³ and in addition, tissue injury due to the large numbers of mitochondria present in heart tissue²⁴. Glutathione (GSH) depletion of cardiomyocytes has also been described as a mechanism of oxidative stress-mediated toxicity of DOX²⁵.

Moreover, the cell death process appears to be preceded by severe deregulation in superoxide anionmediated iron homeostasis and semiquinone form of $DOX^{26,13}$, which reaches the center of the ferritin molecule and promotes the release of ions in the form of Fe²⁺ that as Fe³⁺ rapidly reacts with DOX and gives rise to toxic complexes to cardiomyocytes²⁷.

The myocardial toxicity induced by DOX can be expressed at any stage of chemotherapy, even months or years after its end²⁸. Dilated cardiomyopathy by DOX is usually related to the cumulative dose (> 500 mg per m²) and has an incidence of approximately 1.7% starting from the first month of the last dose of chemotherapy received²⁹.

Part of the toxicity of DOX has been attributed to oxidative stress signaling via cytokines, especially TNF- α , a mediator described as a modulator of cardiac failure in several diseases. It is described that the increased production of reactive species leading to increased expression of TNF- α in cardiomyocytes via activation of NF-kB²², thus perpetuating the inflammatory response in the host through the production of other proinflammatory cytokines such as IL-6³⁰.

Other toxic effects of DOX include acute events as stomatitis, neutropenia, thrombocytopenia, generalized infections, liver abnormalities, hematological toxicity, nausea and myelosuppression. In the long term, some patients may develop leukemia secondary to treatment, necrosis at the injection site, liver injury, reversible alopecia, hyperpigmentation, hypersensitivity and neurotoxicity³¹.

Recent studies have shown that there is the presence of immediate systemic oxidative stress in patients treated with DOX, with oxidative cell damage and development of anemia besides the obvious showed decreased antioxidant capacity due to the drop in levels of reduced glutathione (GSSG) and total plasma antioxidant capacity². It was also demonstrated that treatment with DOX promotes important immunological abnormalities in patients with advanced breast cancer immediately after infusion³², suggesting that the modification of the immune response may be associated with increased oxidative stress and tissue damage also reported during breast cancer chemotherapy with DOX. Some studies affirm that the most serious problem caused by DOX is cardiotoxicity³³ and demonstrate that the loss of cardiac function is commonly a result of this treatment³⁴. As previously described, the production of reactive oxygen species and nitrogen is one of the mechanisms by which DOX exerts its antitumor effect³⁵. Based on this assumption, there are several studies describing the involvement of reactive species produced through the use of DOX as cause of cardiac toxicity in various models. Based on results of electrocardiograms, cardiotoxicity was described as a result of treatment with this anticancer agent, in which transient abnormalities can be observed in ST-T waves, supraventricular tachyarrhythmias and ventricular extrasystoles¹⁹.

Although the efficiency is demonstrated in the early stages of treatment with DOX, its continued infusion may lead to the development of drug resistance, requiring the subsequent use of higher doses to achieve sufficient therapeutic effect³⁴. As already described, oxidative damage to the lung induced by DOX, in the form of dose-dependent lesion in the lung tissue, may be one of the pathogenic factors of pulmonary dysfunction.

To minimize or even avoid the side effects caused by treatment with DOX, numerous strategies have been used, such as administration of the chemotherapeutic agent slowly (6 to 72 hours) using cumulative doses in levels considered safer, periodic monitoring of the cardiac function by supplementary tests (e.g., echocardiography, angiocardiography, endomyocardial biopsy, etc.) and the use of drugs that may act as myocardial protectors (e.g., probucol, dexrazoxane, etc.)^{28,35}.

Experimental and in vitro evidence

Experimental evidence corroborates the relationship between oxidative stress and DOX-mediated toxicity. Evidence from animal studies reveals the involvement of reactive species such as superoxide anion, hydrogen peroxide and hydroxyl radicals^{36,37}. Additionally, the treatment of rat cardiomyocytes in culture with DOX was demonstrated to cause, besides lipid peroxidation, an increase in LDH release and changes in glucose transport to those cells³⁸. Decreased antioxidant activity and increased apoptosis in rat myocardial cells were observed both in vivo³⁹ and in vitro⁴⁰. In rats treated with DOX, in addition to the decrease in cardiac mitochondrial respiration rate there is also an increase in the protein carbonyl content in cardiac tissue⁴¹. A recent study also conducted in rats showed consistent results that pre-treatment with zofenopril can prevent DOX-induced cardiotoxicity⁴¹. Other toxic effects were also observed during experimental chemotherapy with DOX, including nephrotoxicity^{21,39,42,44,45} and lipid peroxidation of pulmonary cells²⁰.

PERSPECTIVES AND CONCLUSIONS

The toxicity of chemotherapy treatment with doxorubicin causes damage to various organs of patients who are in uninterrupted treatment with this antineoplastic agent, therefore causing significant side effects mediated by oxidative stress. Anthracycline-induced cardiotoxicity has been investigated to a great degree and is especially indicated in the literature as the principal side effect. Therefore, care needs to be given to other damage caused by this medication as important as myocardial toxicity, such as renal, pulmonary and liver toxicity, among others. There is a need for further studies to prevent or even encounter a way to control the damage caused by these toxicities in various tissues avoiding future complications.

REFERENCES

- Injac R, Radic N, Govedarica B, Perse M, Cerar A, Djordjevic A, Strukelj B. Acute doxorubicin pulmotoxicity in rats with malignant neoplasm is effectively treated with fullerenol C60(OH)24 through inhibition of oxidative stress. Pharmacol Rep 2009;61:335-42.
- Panis C, Herrera AC, Victorino VJ, Campos FC, Freitas LF, De Rossi T, Colado Simão AN, Cecchini AL, Cecchini R. Oxidative stress and hematological profiles of advanced breast cancer patients subjected to paclitaxel or doxorubicin chemotherapy. Breast Cancer Res Treat 2012;133:89-97.
- Kotamraju S, Konorev EA, Joseph J, Kalyanaraman B. Doxorubicin-induced apoptosis in endothelial cells and cardiomyocytes is ameliorated by nitrone spin traps and ebselen. J Biol Chem 2000;275:33585-92.
- Quiles JL, Huertas JR, Battino M, Mataix J, RamÍrez-Tortosa MC. Antioxidant nutrients and adriamycin toxicity. Toxicology 2002;180:79-95.
- 5. Donkena KV, Young CYF, Tindall DJ. Oxidative stress and dnamethylation in prostate cancer. Obstetr Gynecol Int 2010;2010:1-9.
- Bar-Joseph H, Ben-Aharon I, Tzabari M, Tsarfaty G, Stemmer SM, Shalgi R. In vivo bioimaging as a novel strategy to detectdoxorubicininduced damage to gonadal blood vessels. PLoS ONE 2011;6:1-9.
- 7. Weiss G, Goodnough LT. Anemia of chronic disease. N Engl J Med 2005;352:1011-23.
- Minotti G, Menna P, Salvatorelli E, Cairo G, Gianni L. Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. Pharmacol Rev 2004;56:185-230.
- 9. Gewirtz DA. A critical evaluation of the mechanisms of action proposed for the antitumor effects of the anthracyclin antibiotics adriamycin and daunorubicin. Biochem Pharmacol 1999;57:727-41.
- Casares N, Pequignot MO, Tesniere A, Ghringhelli F, Roux S, Chaput N, Schmitt E, Hamai A, Hervas-Stubbs S, Obeid M, Coutant F, Metivier D, Pichard E, Aucouturier P, Pierron G, Garrido C, Zitvogel L, Kroemer G. Caspase-dependent immunogenicity of doxorubicininduced tumor cell death. J Exp Med 2005;202:1691-701.
- 11. Doroshow JH, Tallent C, Schechter JE. Ultrastructural features of adriamycin-induced skeletal and cardiac muscle toxicity. Am J Pathol 1985;118:288-97.
- 12. Minotti G, Cairo G, Monti E. Role of iron in anthracyclincardiotoxicity: new tunes for an old song? FASEB Journal 1999;113:199-12.
- Xu X, Persson HL, Richardson DR. Molecular pharmacology of the interaction of anthracyclins with iron. Mol Pathol 2005;68(2):261-271.
- Kiyomiya K, Matsuo S, Kurebe M. Differences in intracellular sites of action of Adriamycin in neoplastic and normal differentiated cells. Cancer Chemother Pharmacol 2001;47:51-6.
- Singal, PW, Li T,Kumar D, DanelisenIIIiskovic N. Adriamycin-induced heart failure: mechanism and modulation. Mol Cell Biochemistr 2000;207:77-85.

- Kaiserova H, Den Hartog GJM, Simunek T, Schroterova L, Kvasnickova E, Bast A. Iron is not involved in oxidative stress-mediated cytotoxicity of doxorubicin and bleomycin. Br J Pharmacol 2006;149:920-30.
- Pfizer Laboratories Div Pfizer. Doxorubicin hydrochloride for injection, USP. Avaliable from: <URL:http://dailymed.nlm.nih.gov/ dailymed/archives/fdaDrugInfo.cfm?archiveid=51500> [2012 set 19].
- Halliwell B, Gutteridge JMC. Free radicals in biology and medicine. 4th ed. New York: Oxford University; 2007.
- Mukherjee S, Banerjee SK, Maulik M, Dinda AK, Talwar KK, Maulik SK. Protection against acute adriamycin-induced cardiotoxicity by garlic: role of dogenousantioxidants and inhibition of TNF-alpha expression. BMC Pharmacol 2003;3:16-25.
- 20. Blum RH, Carter SK. Adriamycin: A new anticancer drug with significant clinical activity. Ann Int Med 1974;80:249-59.
- Machado NG, Baldeiras I, Pereira GC, Pereira SP, Oliveira PJ. Subchronic administration of doxorubicin to Wistar rats results in oxidative stress and unaltered apoptotic signaling in the lung. Chemico-Biological Interactions 2010;18:478-86.
- 22. Patel N, Joseph C, Corcoran GB, Ray SD. Silymarin modulates doxorubicin-induced oxidative stress, Bcl-xL and p53 expression while preventing apoptotic and necrotic cell death in the liver. Toxicol Appl Pharmacol 2010;245:143-52.
- 23. Mukherjee S, Banerjee SK, Maulik M, Dinda AK, Talwar KK, Maulik SK. Protection against acute adriamycin-induced cardiotoxicity by garlic: role of endogenous antioxidants and inhibition of TNF-alpha expression. BMC Pharmacol 2003;3:16.
- Doroshow JH, Locker GY, Myers CE. Enzymatic defenses of the mouse heart against reactive oxygen metabolites. J Clin Invest 1980;65:128-35.
- Jackson JA, Reevs JP, Muntz KH, Kruk D, Prough RA, Willerson JT, Buja LM. Evaluation of free radicals effects and catecholamine alterations in adryamicin cardiotoxicity. Am J Pathol 1984;117:140-53.
- Minotti G. Sources and role of iron in lipid peroxidation. Chem Res Toxicol 2003;6:134-46.
- Hasinoff BB, Davey JP. The iron (III) adryamicin complex inhibits cytochrome c oxidase before its inactivation. Biochem J 1988;250:827-34.
- 28. Matos Neto RP, Petrilli AS, Silva CMC, Filho OC, Oporto VM, Gomes LFG, Paiva MG, Carvalho ACC, Moisés VA. Função sistólica do ventrículo esquerdo pela ecocardiografia em crianças e adolescentes com osteossarcoma tratados com doxorrubicina com e sem dexrazoxane. Arq Bras Cardiol 2006;87:763-71.
- Yu PC, Calderaro D, Ikeoka DT, Demarchi LMF, Caramelli B. Toxicidade miocárdica por doxorrubicina. Rev Assoc Med Bras 2005;51:121-32.
- Elsea CR, Roberts DA, Druker BJ, Wood IJ. Inhibition of p38 MAPK suppresses inflammatory cytokine induction by etoposide, 5-fluouracil and doxorubicin, without affecting tumoricidal activity. PLoS ONE 2008;3:e2355.
- 31. Panis C, Lemos LG, Victorino VJ, Herrera AC, Campos FC, Colado Simão AN, Pinge-Filho P, Cecchini AL, Cecchini R. Immunologycal effects of Taxol and Adryamicin in breast cancer patients. Cancer Immunol Immunother 2012;61:481-8.
- 32. Šimùnek T, Štìrba M, Popelová O, Adamcová M, Hrdina R, Geršl V. Anthracycline-induced cardiotoxicity: Overview of studies examining the roles of oxidative stress and free cellular iron. Pharmacol Rep 2009;61:154-71.
- 33. Dayton A, Selvendiran K, Meduru S, Khan M, Kuppusamy ML, Naidu S, Kálai T, Hideg K, Kuppusamy P. Amelioration of doxorubicin-induced cardiotoxicity by an anticancer-antioxidant dual-function compound, HO-3867. J Pharmacol Exp Ther 2011;339:350-7.
- 34. De Rossi T, Panis C, Victorino VJ, Freitas LF, Herrera ACS, Cecchini AL, Cecchini R. Breast cancer and oxidative stress in chemotherapy. Appl Cancer Res 2009;29:150-6.
- 35. Speyer JL, Green MD, Zeleniuch-Jacquotte A, Wernz JC, Rey M, Sanger J, Kramer E, Ferrans V, Hochster H, Meyers M, Blum RH, Feit F, Burrows MA, Muggia FM. ICRF-187 Permits longer treatment with doxorubicin in women with breast cancer. J Clin Oncol 1992;10:117-27.
- 36. Doroshow JH. Effect of Anthracycline antibiotics on oxygen radical formation in rat heart. Cancer Res 1983;43:460-72.

- Doroshow JH, Davies KJA. Redox cycling of anthracyclins by cardiac mitochondria: formation of superoxide anion, hydrogen peroxide and hydroxyl radical. J Biol Chem 1986;261:3068-74.
- 38. Hrelia S, Fiorentini D, Maraldi T, Angeloni C, Bordoni A, Biagi PL, Hakim G. Doxorubicin induces early lipid peroxidation associated with changes in glucose transport in cultures cardiomyocytes. Biochimica Biophysica Acta 2002;1567:150-6.
- 39. Elberry AA, Abdel-Naimb AB, Abdel-Sattar EA, Nagy AA, Mosli HA, Mohamadin AM, Ashour OM. Cranberry (Vaccinium macrocarpon) protects against doxorubicin-induced cardiotoxicity in rats. Food Chem Toxicol 2010;48:1178-84.
- 40. Mukhopadhyay P, Rajesh M, Bátkai S, Kashiwaya Y, Haskó G, Liaudet L, Szabó C, Pacher P. Role of superoxide, nitric oxide, and peroxynitrite in doxorubicin-induced cell death in vivo and in vitro. Am J Physiol Heart Circ Physiol 2009;296:H1466-483.
- Berthiaume JM, Oliveira PJ, Fariss MW, Wallace KB. Dietary vitamin E decreases doxorubicin-induced oxidative stree without preventing mitochondrial dysfunction. Cardiovascular Toxicol 2005;5:271-67.

- Bozcali E, Dedeoglu DB, Karpuz V, Suzer O, Karpuz H. Cardioprotective effects of zofenopril, enalapril and valsartan against ischaemia/ reperfusion injury as well as doxorubicin cardiotoxicity. Acta Cardiol 2012;67:87-96.
- Ayla S,Seckin I, Tanriverdi G, Cengiz M, Eser M, SonerBC, Oktem G.Doxorubicin Induced Nephrotoxicity:Protective Effect of Nicotinamide. Int J Cell Biol 2011;2011:1-9.
- 44. Injac R, Boskovic M, Perse M, Koprivec-Furlan E, Cerar A, Djordjevic A, Strukelj B. Acute doxorubicin nephrotoxicity in rats with malignant neoplasm can be successfully treated with fullerenol C60(OH)24 via suppression of oxidative stress. Pharmacol Rep 2008;60:742-9.
- Mansour MA, El-Kashef HA, Al-Shabanah OA. Effect of captopril on doxorubicin-induced nephrotoxicity in normal rats. Pharmacol Res 1999;39:233-7.